Effect of gold nanoparticles on postoperative peritoneal adhesions in rats

¹A.H. Mohammadpour; ²A. Tavassoli; ³M.R. Khakzad; ⁴E. Zibaee; ⁵M. Afshar; ⁶M. Hashemzaei; ^{7*}Gh. Karimi

¹Department of Clinical Pharmacy, School of Pharmacy, Pharmaceutical Research Center, Mashhad University of Medical Sciences, Mashhad Iran

²Endoscopic & Minimally Invasive Surgery Research Centre, Ghaem hospital, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

³Allergy Research Center, Mashhad University of Medical Sciences, Mashhad, Iran, Department of Immunology, Mashhad Branch, Islamic Azad University, Mashhad, Iran

⁴Department of Clinical Pharmacy, School of Pharmacy, Mashhad University of Medical Science, Mashhad, Iran ⁵Department of Anatomy, Faculty of Medicine, Birjand University of Medical Sciences, Birjand, Iran

⁶Department of Pharmacology and Toxicology, School of Pharmacy, Zabol University of Medical Science, ⁷Medical Toxicology Research Center, School of Pharmacy, Mashhad University of Medical Science,

Mashhad Iran

Received; 17 May 2015 Accepted; 21 June 2015

ABSTRACT:

Objective(s): Abdominal adhesions are one of the most important problems, occurring after intra-abdominal surgery in more than 90% of cases. This condition is the leading cause of bowel obstruction, infertility, and abdominal/pelvic pain. Gold nanoparticles (GNPs) have been shown to be non-toxic and exhibit anti-inflammatory, anti-angiogenic and antioxidant activities. The purpose of this study was to determine the effect of intraperitoneal lavage with GNP solutions on the development of postoperative peritoneal adhesion (PPA).

Materials and Methods: In the current experimental study, thirty-five male Wistar rats were randomly assigned to seven groups of five rats. After a standardized peritoneal injury, GNP solutions in different concentrations (1, 2.5, 5, 10, 50 and 100 ng/ml) were locally administered through nebulization; normal saline (NS) was administered to the control group. Two weeks later, the rats were sacrificed and cecum and peritoneal samples were harvested for histopathological assessment. Blood samples were obtained to determine serum concentrations of inflammatory biomarkers including tumor necrosis factor alpha (TNF-), interleukin-1 beta (IL-1) and vascular endothelial growth factor (VEGF).

Results: The rats treated with GNPs had significantly lower microscopic and macroscopic peritoneal adhesion scores, compared to the control group (P<0.05). Score 5 of macroscopic adhesions was reported in all the rats of the control group, unlike the GNP groups. Furthermore, microscopic adhesions were reported with all rats in the control group, unlike the GNP groups (reported in 0 out of 5 rats in all GNP groups). In addition, serum levels of IL-1, TNF- and VEGF underwent no significant changes.

Conclusion: Compared to the control group, GNPs decreased the severity of peritoneal adhesions, although they did not alter TNF , IL-1 or VEGF serum levels.

Keywords: Gold nanoparticles, Nebulization, Postoperative peritoneal adhesion, Rat

INTRODUCTION

Postoperative peritoneal adhesion (PPA) is a frequent complication after abdominal surgery. In fact, peritoneal injury after abdominopelvic surgery is the most

*Corresponding Author Email: *karimig@mums.ac.ir* Tel: (+98) 38823255 common cause of abdominal adhesion; less common causes include inflammatory conditions, intraperitoneal infections, and abdominal trauma [1]. Fibrous bands between organs or tissues (or both) develop soon after surgery (after about three hours) [2]. Abdominal injuries result in the release of inflammatory cytokines including tumor necrosis factor alpha (TNF-), interleukin-6 (IL-6) and interleukin-1 beta (IL-1)[3, 4].

Note. This manuscript was submitted on May 17, 2015; approved on June 21, 2015

Both TNF, IL-1 are proinflammatory cytokines that play important roles in the early phases of wound healing process and are produced via activated macrophage in peritoneal fluids [5,7]. Furthermore, increased production of IL-1 by activated macrophage induces IL-6 expression [8]. Expression of inducible cyclooxygenase (COX2), as an activated enzyme, leads to increasing levels of inflammatory mediators such as thromboxane, interleukins and prostaglandins [8]. TNF plays an impo-rtant role in adhesion band formation [7, 9]. Recruitment of neutrophils, macrophages and eosinophils leads to the release of fibrinous exudates at the abdominal cavity [2]. Vascular endothelial growth factor (VEGF) is another signal protein, which plays a critical role in the vasculogenesis and angiogenic process [10]. Many previous studies showed that VEGF antibodies reduced the formation of adhesion bands [11-13]. Moreover, significant oxidative stress is induced by the activation of mesothelium and endothelial cells along with the infiltration and activation of neutrophils and macrophages. Ultimately, this inflammatory process leads to the formation of nascent fibrinous adhesions and expands to fibrins [4].Although the application of new surgical techniques such as laparoscopy is expanding, it is not always possible or appropriate to perform all types of surgical interventions for patients. PPA occurs in 93-100% of patients who undergo trans-peritoneal surgery. This condition can induce bowel obstruction, infertility, abdo-minal pain, pelvic pain, and difficulties during the new surgical interventions [9, 11, 14, 15]. Gold in its natural state is a non-toxic compound for humans and is used extensively in cancer diagnosis and treatment. Considering the established use of gold in laboratory and the chemical stability of gold, gold nanoparticles (GNPs) are expected to be harmless [16, 17]. GNP shave been shown to exhibit anti-inflam-matory, immunomodulatory, and antioxidant activities and possess anti-angiogenic, anal-gesic, and restorative effects [8, 18, 19]. In addition, GNPs are widely used in biomedical imaging and diagnostic tests [20]. GNPs pose anti-angiogenic effects via targeting multiple proangiogenic cytokines, which contain heparin-binding domains including VEGF and basic fibroblast growth factor. VEGF has been shown to have wound-healing effects [19]. Given the role of angiogenesis in peritoneal adhesion, this process may be useful for adhesion prevention. Moreover, GNPs decrease TNF , IL-1 levels in synovial fluids of animals with rheumatoid arthritis [21]. Auranofin, a gold-containing sulfur, exhibits antioxidant effects by subsiding cytokine production including IL-6 and IL-1 [8]. IL-6 induces VEGF gene expression, which play a pivotal role in the angiogenesis and is involved in postoperative peritoneal adhesion [8]. The current study is the first to investigate the effects of intra-peritoneal lavage with GNPs on the development of PPA in Wistar rats.

MATERIALS AND METHODS

Materials

GNPs (300 nm in size) were purchased from Sigma-Aldrich (USA) with more than 95% spherical particles. TNF, IL-1 and rat VEGF ELISA kits were purchased from RayBiotech Co. (USA).

Animals

Adult male Wistar rats (weighing 250-300 g) were obtained from the School of Pharmacy of Mashhad University of Medical Sciences. Animals were allocated into groups of five and housed in cages; animals were given food and water ad libitum and kept on a 12-h light/12-h dark cycle. All methods used in this study were in accordance with the guidelines by the declaration of Helsinki for experimental studies on animals [22].

Surgical procedure

Animals were subjected to anesthesia, using an intraperitoneal (IP) injection of ketamine 100 mg/kg and xylazine 10 mg/kg. The abdomen was then shaved and prepared with alcohol and iodine solution. After drying, 3 cm laparotomy was performed to gain access to the abdominal cavity. To form an adhesion, Peritoneal Button Creation (PBC) technique was applied, which is considered the most consistent and reproducible animal technique for the intra-abdominal adhesion model [1]. Barbed 2.0 polypropylene sutures in a chain distribution were used on the parietal peritoneum to create four peritoneal buttons. Each suture encapsulated approx-imately 2 cm of parietal peritoneum; the diameter of the tied sutures were approx-imately 5 mm. Different concentrations of GNPs were locally administered in the peritoneal cavity via nebulization and then the peritoneum was sutured. Two weeks later, animals underwent a laparotomy and cecum and peritoneal samples were sent for histopathological examinations; adhesion formation intensity was recorded according to the Adhesion Scoring System (ASS). Blood samples were obtained to determine the serum concen-trations of inflammatory biomarkers. ELISA method was applied for the determination of serum levels of biomarkers.

Statistical analysis

The results were expressed as mean \pm SD. One-way ANOVA and \div 2 tests were performed, using SPSS version 16 (SPSS Inc., Chicago, Illinois, United States). P-values less than 0.05 were considered statistically significant.

RESULTS

Effects of GNPs on macroscopic results of peritoneal adhesions

All surgical procedures were well tolerated by rats and no infection was observed in animals undergoing surgery. In the control group, severe adhesions were observed between the intestine and peritoneum (P<0.05). In GNP group of 1 ng/ml, one rat had an adhesion, whereas in groups of GNP concentrations of 2.5, 5, 10, 50 and 100 ng/ ml, no adhesions were found; in the control group, adhesions were observed in all the rats, as shown in Fig 1 (P<0.05). All GNP groups showed a significant decrease in severity of adhesion, in comparison with the control group. Error! Reference source not found. represents the adhesion formation and division, based on the Adhesion Scoring System (ASS).

Effects of GNPs on microscopic results of peritoneal adhesions

Histopathological examinations with resp-ect to inflammation, fibrosis and neovas-cularizasion are shown in Fig 2. Microscopic adhesion severity were div-ided into 4 scores based on inflammation, infiltration and tissue fibrosis; score 1; aggregation of fibrin and neutrophil infiltration that is related to the first step of inflammation, score 2; edema, granulation formation, infiltration and migration of connective tissue that is related to proliferation step, score 3; collagen formation and score 4; fibrous formation. All rats showed a significant decrement in microscopic histopathological examina-tions, unlike the control group (P<0.05). As it is shown in Fig 2, score 4 of peritoneal adhesion formation was eported in all rats of the control group. Representative Fig of microscopic findings of hematoxylin and eosin H&E) staining, following the intra-peritoneal administration of GNP solutions, is shown in Error! Reference source not found.



Fig. 1. Comparison of the effects of GNPs on peritoneal macroscopic adhesions in different groups of Wistar rats. Numbers of rats based on adhesion severity scores in GNP (1, 2.5, 5, 10, 50 and 100 ng/ml) and control (NS) groups are shown (**P*< 0.05 was considered significant for each group)



Fig. 2. Comparison of the effects of GNPs on peritoneal microscopic adhesions in different groups of Wistar rats. Numbers of rats based on microscopic findings in GNP (1, 2.5, 5, 10, 50 and 100 ng/ml) and control (NS) groups are shown in the Fig (* P < 0.05 was considered significant)

Effects of GNPs on serum concentration of proinflammatory cytokines (TNF, IL-1) and VEGF

Changes in TNF , IL-1 were compared before and after the interv-ention. There were no significant differences between the GNP groups and the controls or among the GNP groups 14 days after the surgery (Table 1 P>0.05). There were no significant differences among GNP groups and the controls (Table 1. Comparison of changes in TNF , IL-1 serum concentrations in different groups in comparison to control group of Wistar rats., P>0.05). Furthermore, no significant differences were found among the GNP groups (P>0.05).

Gold nanoparticles and postoperative peritoneal adhesions



Fig. 3. Adhesion band scores based on ASS method: A: score 1, single, filmy adhesion, B: score 2, more than one filmy adhesion; C: score 3, single, dense adhesion band; D: score 4 single, dense adhesion band with surface adhesion and score 5, more than one dense adhesion band; A and B are related to GNPs and C and D are related to the NS group



Fig. 4. H&E staining of histological absorption of animal tissues following the administration of GNPs with different concentrations for intraperitoneal lavage. A: fibrilative inflammation and neutrophil infiltration; B: proliferative phase, characterized by the formation of granulation tissues, edema, proliferation, and migration of connective tissue cells; C: scar formation and collagen

DISCUSSION

Our results showed that intra-abdominal lavage with GNPs (300 nm in diameters) in different concentrations significantly reduced PPA formation at microscopic and macroscopic levels, whereas, it did not reduce the levels of proinflammatory biomarkers including TNF , IL-1 and VEGF. GNPs have been shown to have protective

effects on organs without any toxic effects on animal organs including heart, kidney, liver and brain [23]. The role of inflammation and angiogenesis in peritoneal adhesions has been thor-oughly elucidated in previous studies [5, 6, 13, 24, 25]. Following to the injury of peritoneal mesothelium cells, vessels were dilated and

	h TNF-serum concentration (Pre-Post)*	h VEGF serum concentration (Pre- Post)*	h IL-1 serum concentration (Pre-Post)*
Control group	3.42±0.17	3.4±0.02	$4.7 {\pm} 0.02$
GNPs (1%)	3.56 ± 0.32	3.40 ± 0.09	4.72 ± 0.02
GNPs (2.5%)	3.47 ± 0.23	3.49 ± 0.09	4.74 ± 0.05
GNPs (5%)	3.44 ± 0.32	3.50 ± 0.12	4.76 ± 0.09
GNPs (10%)	3.61 ± 0.16	3.39 ± 0.03	4.77 ± 0.08
GNPs (50%)	3.33 ± 0.26	3.39 ± 0.04	4.69 ± 0.02
GNPs (100%)	3.62 ± 0.18	3.41 ± 0.02	4.70 ± 0.08

Table 1. Comparison of changes in TNF-, VEGF and IL-1 serum concentrations in different groups in comparison to control group of Wistar rats

fibrinous exudates infiltrated to the outside of vessels. Fibrous tissue consists of fibroblasts and extracellular matrix that leads to sediment scar formation [26]. After tissue injury, macrophages aggregate in the location and potentiate by inflammatory factors including plasminogen, IL-1, IL-6 and TNF [27] which led to the adhesion of peritoneal site. GNPs showed various anti-inflammatory, anti-oxidant and anti-angiogenic properties and these effects have been mentioned in many different clinical and animal studies [8, 16, 19, 21, 28]. Although the anti-adhesive mechanisms of GNPs are not completely clear, anti-angiogenic and antiinflammatory effects are likely to be the main reasons for GNPs effects. Some drugs such as COX-2 inhibitors, corticosteroids and immunomodulators, have been used to reduce adhesive bands in animal models and have yielded satisfactory results [11, 29, 30]. Our study is in consistence with a study that was done in animal models and pretreated them with selective and nonselective COX inhibitors to prevent the formation of adhesion bands [30]. The results showed that selective and non-selective COX inhibitors significantly reduced adhesion bands [30]. Another study showed that infliximab(a TNF antagonist) with immunomodulatory effects significantly decreased peritoneal adhesion formation in rats [9]. In a study that was conducted on diabetic mice showed that GNPs have free radical scavenging activities and anti-oxidant effects via activation of gluta-thione, superoxide dismutase and catalase [23]. In the aforementioned study, it was confirmed that antioxidant and anti-hyperglycemic effects of GNPs are due to inhibiting of reactive oxygen species (ROS) production or balanced ROS generation in streptozocine-induced hyper-glycemic mice; this inhibition showed antioxidant and free radical scavenging effects via increasing levels of antioxidant defense enzymes [23]. Previous studies showed that intra-peritoneal administration of antioxidants posed

anti-adhesive effects [15, 31]. Intra-peritoneal administration of honey and aloe vera gel with antioxidant effects decreased intra-abdominal adhesions [15, 32, 31]. Taken together, the current study showed that GNPs decreased adhesive bands following local administration in peritoneal area but did not reduce serum levels of TNF, IL-1 or VEGF in the GNPs treated group. Explanation for the observed results might includ a) given the local administration of GNPs (instead of its systemic application), local content (peritoneal exudates content) level of biomarkers need to be evaluated. If GNPs were administered intravenously, serum levels of mediators such as TNF

, IL-1 and VEGF might have significantly changed. Cheong and colleagues showed that peritoneal exudate concentrations of cytokines are different from their serum levels [7]. They showed that adhesion formation within 12h after surgery was correlated with IL-1 and IL-6 intra-peritoneal concentration [7]; b) other adhesive, angiogenic and pro-inflammatory factors in the formation of postoperative adhesion bands may have a role; c) significant changes in the serum levels of biomarkers depends on the pathophysiology of adhesion bands due to the involvement of many factors in their formation ; d) if the diameters of GNPs had changed, perhaps, the results would have altered, as well.

CONCLUSION

In the present study, it was confirmed that GNPs (300 nm) decreased the formation of postoperative adhesive bands at micros-copic and macroscopic levels. However, no decrement was observed in plasma levels of TNF, IL-1 or VEGF.

ACKNOWLEDGMENTS

The authors thank the Vice Chancellor of Research, Mashhad University of Medical Sciences for financial support.

REFERENCES

- Whang SH, Astudillo JA, Sporn E, Bachman SL, Miedema BW, Davis W, et al. In Search of the Best Peritoneal Adhesion Model: Comparison of Different Techniques in a Rat Model. J Surg Res. 2011; 167(2): 245-250.
- 2.Jomezadeh V, Mohammadpour AH, Rajabi O, Tavassoli A, Maddah G. Evaluation of curcumin effects on post-operative peritoneal adhesion in rats. Iran J Basic Med Sci. 2012; 15(6): 1162-7.
- Cheong YC, Laird SM, Shelton JB, Ledger WL, Li TC, Cooke ID. The correlation of adhesions and peritoneal fluid cytokine concentrations: a pilot study. Hum Reprod. 2002; 17(4): 1039-1045.
- 4.Liakakos T, Thomakos N, Fine PM, Dervenis C, Young RL Peritoneal adhesions: etiology, pathophysiology, and clinical significance. Dig. sur., 2001; 18(4): 260-273.
- Tsukada K, Katoh H, Shiojima M, Suzuki T, Takenoshita S, Nagamachi Y. Concentrations of cytokines in peritoneal fluid after abdominal surgery. Eur J Surg. 1993; 159(9): 475-9.
- 6.Kaidi AA, Nazzal M, Gurchumelidze T, Ali MA, Dawe EJ, Silva YJ. Preoperative administration of antibodies against tumor necrosis factor-alpha (TNF-alpha) and interleukin-1 (IL-1) and their impact on peritoneal adhesion formation. Am Surg. 1995; 61(7): 569-72.
- 7.Cheong YC, Laird SM, Shelton JB, Ledger WL, Li TC, Cooke ID. The correlation of adhesions and peritoneal fluid cytokine concentrations: a pilot study. Hum Reprod. 2002; 17(4): 1039-1045.
- Kim NH, Lee MY, Park SJ, Choi JS, Oh MK, Kim IS. Auranofin blocks interleukin 6 signalling by inhibiting phosphorylation of JAK1 and STAT3. Immunology. 2007; 122(4): 607-614.
- Kurukahvecioglu O, Koksal H, Gulbahar O, Erdem O, Engin D, Yazicioglu O, et al. Infliximab" TNF-alpha antagonist" decreases intraabdominal adhesions. Saudi Med. J., 2007; 28(12): 1830-1835.
- 10.Yoshida A, Anand-apte B, Zetter BR. Differential Endothelial Migration and Proliferation to Basic Fibroblast Growth Factor and Vascular Endothelial Growth Factor. Growth Factors.1996; 13(1-2): 57-64.
- 11.Saltzman AK, Olson TA, Mohanraj D, Carson LF, Ramakrishnan S. Prevention of postoperative adhesions by an antibody to vascular permeability factor/vascular endothelial growth factor in a murine model. Am J Obstet Gynecol, 1996; 174(5): 1502-6.
- 12.Diamond MP, El-Hammady E, Munkarah A, Bieber EJ, Saed G Modulation of the expression of vascular endothelial growth factor in human fibroblasts. Fertil Steril. 2005; 83(2): 405-9.
- Cahill RA, Redmond HP. Cytokine orchestration in postoperative peritoneal adhesion formation. World J Gastroenterol. 2008; 14(31): 4861-4866.
- 14.Zheng Z, Zhang W, Sun W, Li X, Duan J, Cui J, et al. Influence of the carboxymethyl chitosan anti-adhesion solution on the TGF-â1 in a postoperative peritoneal adhesion rat. J Mater Sci Mater Med. 2013; 24(11): 2549-2559.
- 15.Yuzbasioglu MF, Kurutas EB, Bulbuloglu E, Goksu M, Atli Y, Bakan V, et al. Administration of honey to prevent peritoneal adhesions in a rat peritonitis model. Int J Surg. 2009; 7(1): 54-57.

- 16.Pal R, Panigrahi S, Bhattacharyya D, Chakraborti AS. Characterization of citrate capped gold nanoparticlequercetin complex: Experimental and quantum chemical approach. J Mol Struct. 2013; 1046(0): 153-163.
- 17.Kalishwaralal K, Deepak V, Ram Kumar Pandian S, Gurunathan S. Biological synthesis of gold nanocubes from Bacillus licheniformis. Bioresour Technol. 2009; 100(21): 5356-5358.
- 18.Norton S. A brief history of potable gold. Vol. 8. 2008; 120-123.
- 19.Arvizo RR, Rana S, Miranda OR, Bhattacharya R, Rotello VM, Mukherjee P. Mechanism of anti-angiogenic property of gold nanoparticles: role of nanoparticle size and surface charge. Nanomedicine. 2011; 7(5): 580-587.
- 20.Dykman LA, Khlebtsov NG. Gold nanoparticles in biology and medicine: recent advances and prospects. Acta Naturae. 2011; 3(2): 34-55.
- 21.Tsai C-Y, Shiau A-L, Chen S-Y, Chen Y-H, Cheng P-C, Chang M-Y, et al. Amelioration of collagen-induced arthritis in rats by nanogold. Arthritis Rheum., 2007; 56(2): 544-554.
- 22.Carlson RV, Boyd KM, Webb DJ. The revision of the Declaration of Helsinki: past, present and future. Br J Clin Pharmacol. 2004; 57(6): 695-713.
- 23.BarathManiKanth S, Kalishwaralal K, Sriram M, Pandian S, Youn H-s, Eom S, et al. Anti-oxidant effect of gold nanoparticles restrains hyperglycemic conditions in diabetic mice. J Nanobiotechnology. 2010; 8(1): 1-15.
- 24.diZerega GS.Biochemical events in peritoneal tissue repair. Eur J Surg Suppl. 1997; (577): 10-16.
- Thornton FJ, Barbul A.Healing in the gastrointestinal tract. Surg Clin North Am. 1997; 77(3): 549-573.
- 26.Eckes B, Nischt R, Krieg T.Cell-matrix interactions in dermal repair and scarring. Fibrogenesis & Tissue Repair. 2010; 3: 4-4.
- Vykoukal D, Davies MG.Vascular biology of metabolic syndrome. J Vasc Surg. 2011; 54(3): 819-831.
- Shah ZA, Vohora SB. Antioxidant/Restorative Effects of Calcined Gold Preparations Used in Indian Systems of Medicine against Global and Focal Models of Ischaemia. Pharmacology & Toxicology, 2002; 90(5): 254-259.
 Saed GM, Munkarah AR, Diamond MP.Diamond,
- 29.Saed GM, Munkarah AR, Diamond MP.Diamond, Cyclooxygenase-2 is expressed in human fibroblasts isolated from intraperitoneal adhesions but not from normal peritoneal tissues. Fertil Steril. 2003; 79(6): 1404-1408.
- 30.Greene AK, Alwayn IP, Nose V, Flynn E, Sampson D, Zurakowski D, et al.Prevention of intra-abdominal adhesions using the antiangiogenic COX-2 inhibitor celecoxib. Ann Surg. 2005; 242(1): 140.
 31.Aysan E, Bektas H, Ersoz F.A new approach to
- 31.Aysan E, Bektas H, Ersoz F.A new approach to postoperative peritoneal adhesions: prevention of peritoneal trauma by aloe vera gel. Eur J Obstet Gynecol Reprod Biol. 2010; 149(2): 195-198.
- 32.Pereira DV, Petronilho F, Pereira HR, Vuolo F, Mina F, Possato JC, et al.Effects of gold nanoparticles on endotoxininduced uveitis in rats. Invest Ophthalmol Vis Sci. 2012; 53(13): 8036-8041.

How to cite this article:

Mohammadpour AH, Tavassoli A, Khakzad MR, Zibaee E, Afshar M, Hashemzaei M, Karimi Gh. Effect of gold nanoparticles on postoperative peritoneal adhesions in rats. Nanomed. J., 2015; 2(3): 211-216.